

·综述·

外泌体在妇科恶性肿瘤中的作用及临床应用进展

张凤梅¹, 李红芳²

(1. 甘肃中医药大学第一临床医学院, 甘肃 兰州 730000; 2. 兰州市第一人民医院妇产科, 甘肃 兰州 730000)

摘要: 外泌体是具有脂质双层膜结构的小囊泡, 因其非侵入性、高可及性和稳定性应用于精准医学中, 在肿瘤的转移、侵袭和血管生成等过程中发挥重要作用。妇科恶性肿瘤主要包括宫颈癌、卵巢癌和子宫内膜癌, 其早期诊断和治疗一直是研究的热点和难点。外泌体作为新型的生物学标志物, 具有高度特异性, 能够有效阻断妇科恶性肿瘤的发生和发展。本文详细探讨了外泌体在宫颈癌、卵巢癌和子宫内膜癌中的诊断和治疗应用。在宫颈癌中, 外泌体参与HPV感染、血管生成和免疫逃逸等过程, 其携带的特定miRNA(如miR-30d-5p、let-7d-3p)可作为诊断标志物。此外, 外泌体还可作为靶向药物递送载体和疫苗研发平台。在卵巢癌中, 外泌体携带的miRNA(如miR-21、miR-200家族)对早期诊断具有参考价值, 且外泌体在化疗耐药和肿瘤进展中扮演重要角色。对于子宫内膜癌, 外泌体中的miRNA(如miR-15a-5p、miR-106b-5p)可作为早期检测的生物标志物。此外, 本文还指出了外泌体在临床应用面临的挑战, 如分离提取的复杂性和细胞来源的识别等, 并强调了进一步的基础研究和临床试验的必要性。本文为妇科恶性肿瘤的早期诊断和精准治疗提供了新的思路和方法, 具有重要的理论和临床意义。

关键词: 外泌体; 妇科恶性肿瘤; 宫颈癌; 卵巢癌; 子宫内膜癌; 临床应用

中图分类号: R737.3

文献标志码: A

文章编号: 1672-3554(2025)02-0266-09

DOI: 10.13471/j.cnki.j.sun.yat-sen.univ(med.sci).2025.0210

The Role and Clinical Application Progress of Exosomes in Gynecological Malignancies

ZHANG Fengmei¹, LI Hongfang²

(1. First Clinical Medical School, Gansu University of Chinese Medicine, Lanzhou 730000, China; 2. Department of Obstetrics and Gynecology, The First People's Hospital of Lanzhou City, Lanzhou 730000, China)

Correspondence to: LI Hongfang; E-mail: 623830138@qq.com

Abstract: Exosomes are small vesicles with a lipid bilayer membrane structure that have applied in precision medicine due to their non-invasive nature, high accessibility, and stability. Exosomes play a crucial role in processes such as tumor metastasis, invasion, and angiogenesis. Gynecological malignancies primarily include cervical cancer, ovarian cancer, and endometrial cancer, and their early diagnosis and treatment have long been a focus of research. As novel biological markers, exosomes exhibit high specificity and can effectively block the occurrence and progression of gynecological malignancies. This article explores the diagnostic and therapeutic applications of exosomes in cervical cancer, ovarian cancer, and endometrial cancer in detail. In cervical cancer, exosomes are involved in processes such as HPV infection, angiogenesis, and immune evasion, with specific miRNAs (such as miR-30d-5p and let-7d-3p) serving as diagnostic markers. Furthermore, exosomes can act as targeted drug delivery vehicles and vaccine development platforms. In ovarian cancer, the miRNAs carried by exosomes (such as miR-21 and the miR-200 family) have reference value for early diagnosis, and exosomes play an important role in chemotherapy resistance and tumor progression. For endometrial cancer, miRNAs in exosomes (such as miR-15a-5p and miR-106b-5p) can serve as biomarkers for early

收稿日期: 2024-11-20

录用日期: 2025-02-23

基金项目: 兰州市卫生健康行业科研项目(A2023008)

作者简介: 张凤梅, 第一作者, 研究方向: 妇科肿瘤, E-mail: 942519326@qq.com; 李红芳, 硕士生导师, 主任医师, 研究方向: 妇科肿瘤, E-mail: 623830138@qq.com

detection. Additionally, this article highlights the challenges faced by exosomes in clinical applications, such as the complexity of isolation and extraction and the identification of cell sources, and emphasizes the necessity for further basic research and clinical trials. This study provides new ideas and methods for the early diagnosis and precision treatment of gynecological malignancies, holding significant theoretical and clinical importance.

Key words: exosomes; gynecological malignancies; cervical cancer; ovarian cancer; endometrial cancer; clinical application

[J SUN Yat-sen Univ (Med Sci), 2025, 46(2): 266-274]

妇科恶性肿瘤是女性中较为常见的疾病之一,预防对限制其发病率和死亡率至关重要。但目前并非所有恶性肿瘤都有有效的筛查方法,因此医学界正开发新的生物标志物和方法来进行及时有效的早期诊断及治疗。最常见的妇科恶性肿瘤包括宫颈癌、卵巢癌和子宫内膜癌。2022年全球癌症统计数据,宫颈癌发病率排名第8位,全球有661 021例新发病例,死亡率排名第9位,有348 189例死亡病例;卵巢癌发病率排名第18位,全球有324 398例新发病例,死亡率排名第14位,有206 839例死亡病例;子宫内膜癌发病率排名第15位,全球有420 242例新发病例,死亡率排名第19位,有97 704例死亡病例^[1]。预防妇科恶性肿瘤是降低女性生殖系统患癌症风险的根本途径^[2]。新型生物学标志物的发现可以有效预防癌症的发生。外泌体标志物在恶性肿瘤中明显升高,具有高度特异性,因此外泌体的研究有助于癌症的早期发现与预防^[3]。

1 外泌体概述

外泌体最早由 Bonucci 和 Anderson 在 1960 年代后期首次报道,随后发现它们存在于绵羊网织红细胞中^[4]。外泌体是由多种细胞类型产生的细胞外囊泡,包括免疫细胞(巨噬细胞、NK 细胞、树突状细胞、T 细胞、B 细胞)、干细胞和癌细胞,其结构与细胞相似,呈凹面半球形,直径范围为 30~150 nm^[5]。发现初期,它们的作用被忽视,随着分子生物学的发展,现发现外泌体具有高含量细胞骨架蛋白、MHCI 类和 II 类蛋白、黏附蛋白(如四聚体、整合素)以及包封的核酸,携带 DNA、RNA、蛋白质等重要物质,充当细胞间通讯调节因子^[6],促进细胞间的物质交换及信息传递。细胞的来源决定了它们的功能和组成成分^[7],因此,明确外泌体的细胞来

源是近年来的研究热点及难点。目前,国内外的研究均表明,外泌体在肿瘤领域扮演着至关重要的角色,它们不仅参与肿瘤的形成,还在其发展的多个关键阶段发挥着重要作用^[8]。

2 外泌体与宫颈癌

宫颈癌是严重威胁女性健康疾病之一^[9],是女性常见的生殖系统疾病,其发病率远超于卵巢癌、子宫内膜癌。最新数据显示,宫颈癌在全球女性癌症中发病率与死亡率均排名第四位^[10],在中国,新发病例和死亡病例为 15.1 万例和 5.6 万例,标化发病率和标化死亡率世界排名分别位于第 95 和 126 位。据估计,2020 年有 100 万儿童成为孤儿,因他们的母亲均死于癌症,其中有近一半孤儿是因母亲死于宫颈癌的结果。近几年来,宫颈癌逐渐呈现出年轻化趋势,虽然总体发病率逐渐减少^[11-12],存活率仍然很低,在癌症中仍位居前列。据估计,到 2030 年,全球宫颈癌病例数量将增加约 50%^[13]。因此宫颈癌的早期诊断显得格外重要。

2.1 外泌体在宫颈癌诊断中的作用与应用

宫颈癌是病毒感染性疾病,主要以人乳头瘤病毒(human papillomavirus, HPV)持续感染为主,临床主要的检测方法也是主要针对 HPV 为主,前期筛查主要是高危型 HPV 和/或新柏氏液基细胞学检测,因经济、地域、文化水平等影响因素^[14],筛检率较低且无性生活者不建议检测,筛检阳性者需进一步行阴道镜检查,必要者取宫颈活检组织,进行病理学检查,患者体验感差,且易感染及出血,不免造成心理阴影。液体活检是一种无创、准确性高和易操作方式,具有明显的独特性,可以追踪癌症的进展,具有症状出现前检测出癌症的优势^[15]。外泌体是液体活检的关键对象之一。Peng 等^[16]研究发现,外泌体参与宫颈癌的 HPV 感染、血管生成、免疫逃

逸及癌细胞的增殖、侵袭过程^[17-21]。此外,外泌体在宫颈癌的治疗领域展现出巨大的应用前景,如靶向药物的输运、疫苗的研发以及液体活检生物标志物等^[22]。因外泌体自身结构的独特性,可成为靶向药物递送的理想载体^[23]。Liang^[24]等发现载有紫杉醇(paclitaxel, PTX)的AS1411-chol外泌体(通过化学修饰的生物纳米载体)可以被有效地递送至靶癌细胞,为癌症治疗提供有前途的递送平台。Hao等^[25-26]研究团队在DC疫苗(sipuleucel-T)的研发方面,进一步用外泌体为载体负载HPV16E7蛋白作为宫颈癌疫苗的研发。Zheng等^[27]对121份血浆样本(23例对照样本、5例CINI、59例CINII-III、21例宫颈鳞状细胞癌和13例腺癌)进行了外泌体miRNA的研究,通过测序、qRT-PCR、ddPCR等方法,得出结论,外泌体miRNA-30d-5p和let-7d-3p是宫颈癌有价值的诊断生物标志物。目前大多数外泌体是从细胞培养上清液和复杂的生物体液(如血浆)中分离出来的,因此,外泌体的产量和纯度是有限的。当使用外泌体作为免疫疗法或其他方法时,必须实现大规模稳定的制备方法^[28-30]。外泌体在如何进一步提升其靶向性、提高药物载装效率以及控制药物释放都是亟须解决的技术瓶颈。目前尚缺乏大规模临床试验的数据支持其安全性和疗效。尽管一些研究报道了外泌体大规模生产和生物相容性改进的方案,但还需要进一步的临床前和临床研究进行验证。

2.2 外泌体在宫颈癌靶向治疗中的作用

外泌体miRNA在宫颈癌的靶向治疗方面也表现出独有的优势性,因自身具有独特的结构及稳定性^[31-34]。Zhu等^[35]研究癌症来源的外泌体miR-651抑制了顺铂耐药性并直接靶向自噬相关基因3 (autophagy-related 3, ATG3),验证外泌体miR-651可能是一种治疗剂,用于寻找预测化疗敏感性的特定治疗靶点。Konishi等^[36]证实外泌体miR-22可能成为宫颈癌放射治疗的一种有前途的药物递送系统。

还有研究发现在早期进展的宫颈癌患者中,miRNA-1228-5p、miRNA-33a-5p、miRNA-3200-3p和miRNA-6815-5p水平降低,miRNA-146a-3p水平升高,而在中晚期miRNA-605-5p、miRNA-6791-5p、miRNA-6780a-5p和miRNA-6826-5p水平升高和miRNA-16-1-3p(或15a-3p)水平降低,因此可以作为宫颈癌预后的检测指标^[37]。外泌体在宫颈癌诊断及治疗中的作用和临床应用详见表1^[38-49]。

3 外泌体与卵巢癌

卵巢癌在全球女性癌症中发病率与死亡率均排名第8位,是妇科恶性肿瘤相关的死亡的主要原因^[50]。任何年龄段均可发病,大多数隐匿性发作,通常诊断出来即是晚期,恶性程度极高,并且与预后不良有关^[51]。尽管卵巢癌的诊治方法日趋成熟,70%的患者在确诊时已属于晚期,5年生存率不足50%^[52]。卵巢癌作为一种常见的妇科肿瘤,有许多不同的组织类型。其中90%为上皮细胞型,其余为非上皮性卵巢癌。还有一些罕见的病理类型,如小细胞癌和癌肉瘤。不同组织类型的卵巢癌具有不同的分子变化、临床行为和治疗效果^[53-54]。

3.1 外泌体在卵巢癌诊断中的作用与应用

因卵巢恶性肿瘤自身独特性,发病年龄广、病情隐匿、盆腹腔位置深,造成了极高的死亡率,预后极差^[55]。现临床上关于卵巢癌的诊断方法,主要是影像学检查和肿瘤标志物检测(如AFP、CEA、CA199、CA125、HE4),特异性低,不灵敏^[56],因此急需新型标志物的出现。外泌体结构稳定,广泛存在于血液、腹水、脑脊液等体液中,可成为卵巢癌未来诊断、治疗和预后的生物标志物之一^[57]。Ho等^[58]研究发现,VI型胶原 $\alpha 3$ (collagen type VI alpha 3, COL6A3)在促进肿瘤进展和转移中至关重要,可作为一种有价值的生物标志物,用于化疗耐药、转移、复发的早期诊断,以及通过检查上皮性卵巢癌(epithelial ovarian cancer, EOC)患者血液、腹水或组织中COL6A3外泌体来预测生存结局,探索可能相关的信号介导,在未来有望为EOC患者开发治疗靶向COL6的偶联抗体或疫苗。卵巢癌细胞衍生的外泌体被证明对卵巢癌的早期诊断有很高的参考价值,研究证实,miRNA-21、miRNA-141、miRNA-200a、miRNA-200b、miRNA-200c、miRNA-203在卵巢癌外泌体中明显升高^[59]。

3.2 外泌体在卵巢癌治疗中的作用与应用

卵巢癌的治疗也是难点,早期主要以手术为主,晚期主要是肿瘤细胞减灭术和联合化疗,以及近年来靶向和免疫治疗,大部分患者经过规范化治疗可获得缓解,但卵巢癌复发率较高,70%患者在2~3年内存在复发情况。因此探索新的治疗方法是近年来卵巢癌研究的热点和难点^[60]。Cao等^[61]发现miR-30a-5p过表达可显著增强HO8910和

表1 外泌体在宫颈癌诊断及治疗中的作用及临床应用

Table 1 The role and clinical application of exosomes in the diagnosis and treatment of cervical cancer

Content	Molecules	Function	Targets	Expression level	Clinical value	References
circRNA	CircSLC26A4	cancer-causing gene	/	upregulated	Biomarker	[38]
miRNA	miR-30d-5p	inhibit cervical cancer cells	/	upregulated	diagnosis/treatment	[39]
miRNA	miR-30a-5p	tumor suppressor	/	upregulated	treatment	[40]
miRNA	miRNA-10a-5p	tumor suppressor	UBE2I	upregulated	treatment	[41]
lncRNA	lncRNA TINCR	oncogene	miRNA-7/ mTOR	knockdown	prognosis	[42]
microRNAs	miR-21	tumor suppressor gene	RECK	upregulated	therapy	[43]
microRNAs	miR-379/miR-656	tumor suppressor.	C14MC	downregulation	diagnostic and prognostic	[44]
miRNA	miR-195-5p	cancer suppressor gene	ATG9A	upregulated	targeted therapy	[45]
circRNA	circ_0039787	oncogene	miR-877- 5p/ KRAS	knockdown	biomarker/therapeutic target	[46]
lncRNAs	LINC00324	oncogene	miR-195-5p	upregulated	prognostic molecular marker	[47]
miRNA	miR-34b-3p	tumor suppressor	STC2/ FN1	upregulated	therapeutic	[48]
miRNA	miR-141	oncogene	Ceacam/ KLRC	upregulated	therapeutic target	[49]

HO8910PM 细胞的增殖、迁移和侵袭能力,而 miR-30a-5p 抑制则相反(P 均 <0.05)。此外,miR-30a-5p 可能通过上调 ZEB2 和 LDH2 参与这些致癌过程,miRNA-30a-5p 促进了卵巢癌细胞的恶性行为,这可能作为 OC 患者的一种有前途的诊断和预后标志物。Wang 等^[62]研究发现 circRAD23B 通过吸附 miR-1287-5p 消除了其对 Y-box 结合蛋白 1 的抑制作用,促进卵巢癌的进展并诱导细胞卡铂耐药,结论 circRAD23B 在卵巢癌细胞的细胞质中稳定表达,在耐药组织中显著增加,并与不良预后相关。外泌体在卵巢癌诊断及治疗中的作用及临床应用详见表 2^[63-69]。

4 外泌体与子宫内膜癌

子宫内膜癌是雌激素密切相关的妇科肿瘤^[70],在全球女性癌症中发病率排名第 6 位,虽然 67% 的患者表现为早期疾病,5 年总生存率(overall survival, OS)为 81%,但 IVA 期和 IVB 期 EC 的 5 年

OS 分别仅为 17% 和 15%^[71]。临床症状不典型,无特异性标志物。因此需要新型标志物的出现,提高早期诊断及治疗。

Zhou 等^[72]研究人员发现,通过候选 miRNA 液滴数字 PCR (droplet digital PCR, ddPCR) 在 202 份独立血浆样本中进一步验证,通过 qRT-PCR 在 32 对子宫内膜肿瘤和邻近正常组织中进行验证,并通过 ddPCR 对 12 例患者术前和术后匹配的血浆样本进行验证。与健康受试者相比,从子宫内膜癌患者血浆样本中分离的外显子组中 miR-15a-5p、miR-106b-5p 和 miR107 显著上调。特别是,单独的 miR-15a-5p 产生的 AUC 值为 0.813,以区分 I 期子宫内膜癌患者和健康受试者。miR-15a-5p 与血清肿瘤标志物(CEA 和 CA125)的整合达到了更高的 AUC 值 0.899。因此 miR-15a-5p 可作为子宫内膜癌早期检测的有前途的诊断生物标志物。miR-142-3p、miR-146a-5p 和 miR-151a-5p 在子宫内膜癌患者血浆中显著过表达^[73]。miR-143-3p、miR-195-5p、miR-20b-5p、miR-204-5p、miR-423-3p 和

表2 外泌体在卵巢癌诊断及治疗中的作用及临床应用

Table 2 The role and clinical application of exosomes in the diagnosis and treatment of ovarian cancer

Content	Molecules	Function	Targets	Expression level	Clinical value	References
lncRNA	SNHG17	carcinogenesis	FOXA1/ miR-485-5p/ AKT1	upregulated	biomarker and therapeutic target	[63]
circRNA	circTMC03	oncogene	miR-515-5p/ITGA8	upregulated	therapeutic	[64]
Protein	USP43	carcinogenic	HDAC2	upregulated	prognosis	[65]
miRNA	miR-223	tumor suppressor	/	upregulated	diagnostic	[66]
miRNA	miR-296-3p	tumor suppressor	PTEN/AKT/SOCS6/STAT3	upregulated	biomarker and therapeutic target	[67]
lncRNA	LncRNA IL21-AS1	oncogene	CD24	upregulated	therapeutic	[68]
circRNAs	circDENND4C	tumor suppressor	miR-200b/c	downregulation	tumor marker	[69]

miR-484在EC患者血清中显著过表达^[74]。此外,来自子宫内膜癌患者尿液来源的外泌体显示miR-200c-3p显著升高^[75]。miR-133a在子宫内膜癌细胞和源自子宫内膜癌细胞的外泌体中高度表达,可

被正常子宫内膜细胞吸收^[76]。外泌体在子宫内膜癌诊断及治疗中的作用及临床应用详见表3^[77-82]。

表3 外泌体在子宫内膜癌诊断及治疗中的作用及临床应用

Table 3 The role and clinical application of exosomes in the diagnosis and treatment of endometrial cancer

Content	Molecules	Function	Targets	Expression level	Clinical value	References
lncRNA	lncRNA DLEU1	oncogenes	microRNA-E2F3	upregulated	therapeutic target	[77]
miRNA	miR-26a-5p	oncogenes	/	downexpression	treatment	[78]
circRNA	hsa_circ_0001610	/	miR-139-5p/ B1	overexpression	target	[79]
lncRNA	lncRNA NEAT1	regulators	miR-26a/b-5p-STAT3-YKL-40	up-regulation	treating	[80]
lncRNA	LncRNA FIRRE	/	miR-199b-5p/SIRT1/BECN1	up-regulation	treatment	[81]
lncRNA	LncRNA TRPM2-AS	/	miR-497-5p/SPP1	upregulated	treatment.	[82]

5 总结与展望

外泌体是一种新型的癌症生物标志物来源,因为它们:①从活的癌细胞中主动分泌;②包含有关肿瘤状况的信息;③可以很容易地从生物体液中提取;④提供易于从高丰度蛋白质中提取的信息;⑤非常稳定。重要的是,外泌体是由活跃增殖的肿瘤细胞分泌的,与凋亡细胞体不同^[83]。外泌体携带细

胞类型特异性RNA和蛋白质分子,它们还可以揭示很多关于肿瘤标志的信息。已确定外泌体包含许多不同的细胞表面受体和蛋白质(热休克蛋白、黏附分子、细胞骨架蛋白以及与融合和膜运动相关的蛋白质)。此外,外泌体含有miRNA分子,这可能会影响供体细胞和受体细胞之间的相互作用。但是,外泌体在实际临床操作中仍存在可能面临的挑战:外泌体的标准分离、提取耗时,对技术及

仪器设备条件要求较高,因外泌体来源于细胞,具体细胞来源难以识别,目前临床应用仍然处于探索阶段,只有少量外泌体基础的治疗方法进入临床试验,如本文2.1提到靶向药物的运输^[84]、疫苗的研发^[85]、液体活检生物标志物^[86]等治疗方法进入临床试验阶段,如治疗妇科恶性肿瘤小分子或核酸药物,可以掺入外泌体中,然后递送到特

定类型的细胞或组织中,实现靶向药物递送,在疫苗研发方面对外泌体进行工程化修饰,增加在肿瘤免疫治疗中的应用^[87-88]。目前尚不完全清楚外泌体是如何促进癌症的发展和转移。因此关于外泌体在妇科恶性肿瘤中如何具体的影响肿瘤分子机制,还需要我们大量基础研究及临床试验。

参考文献

- [1] Bray F, Laversanne M, Sung H, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries [J]. *CA Cancer J Clin*, 2024, 74(3): 229-263.
- [2] Ferrari F, Giannini A. Approaches to prevention of gynecological malignancies [J]. *BMC Womens Health*, 2024, 24(1): 254.
- [3] Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries [J]. *CA Cancer J Clin*, 2021, 71(3): 209-249.
- [4] Johnstone RM, Adam M, Hammond JR, et al. Vesicle formation during reticulocyte maturation. association of plasma membrane activities with released vesicles (exosomes) [J]. *J Biol Chem*, 1987, 262(19): 9412-9420.
- [5] 陈韩, 杨江怡, 陈华, 等. 血清外泌体 let-7d-3p 与 miR-30d-5p 在宫颈癌中的诊断价值 [J]. *中国优生与遗传杂志*, 2023, 31(7): 1366-1370.
Chen H, Yang JY, Chen H, et al. Diagnostic value of serum exosome let-7d-3p and miR-30d-5p in cervical cancer [J]. *Chin J Birth Heal Hered*, 2023, 31(7): 1366-1370.
- [6] Yang C, Huang T, Liang Y, et al. CTHRC1 targeted by miR-30a-5p regulates cell adhesion, invasion and migration in lung adenocarcinoma [J]. *J Cardiothorac Surg*, 2022, 17(1): 46.
- [7] Kok VC, Yu CC. Cancer-derived exosomes: their role in cancer biology and biomarker development [J]. *Int J Nanomedicine*, 2020, 15: 8019-8036.
- [8] 张隆泽, 汪显耀, 何志旭. 外泌体来源的环状RNA对肿瘤细胞和免疫细胞的调控作用研究进展 [J]. *遵义医科大学学报*, 2024, 47(5): 522-530.
Zhang LZ, Wang XY, He ZX. Progress of exosome-derived circular RNA regulation on tumor and immune cells [J]. *J Zunyi Med Univ*, 2024, 47(5): 522-530.
- [9] Olusola P, Banerjee HN, Philley JV, et al. Human Papilloma virus-associated cervical cancer and health disparities [J]. *Cells*, 2019, 8(6): 622.
- [10] 姚一菲, 孙可欣, 郑荣寿. 《2022全球癌症统计报告》解读: 中国与全球对比 [J]. *中国普外基础与临床杂志*, 2024, 31(7): 769-780.
- [11] Yao YF, Sun KX, Zheng RS, et al. Interpretation and analysis of the global cancer statistics report 2022: a comparison between China and the world [J]. *Chin J Bas Clin Gen Surg*, 2024, 31(7): 769-780.
- [12] Zhetpisbayeva I, Kassymbekova F, Sarmuldayeva S, et al. Cervical cancer prevention in rural areas [J]. *Ann Glob Health*, 2023, 89(1): 75.
- [13] 葛尧, 周源, 李鸿博, 等. 外泌体及外泌体 miRNA 在宫颈癌中的研究进展 [J]. *现代肿瘤医学*, 2022, 30(9): 1710-1715.
Ge Y, Zhou Y, Li HB, et al. Research progress of exosome and exosomal miRNA in cervical cancer [J]. *J Mod Oncol*, 2022, 30(9): 1710-1715.
- [14] Ferlay J, Colombet M, Soerjomataram I, et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods [J]. *Int J Cancer*, 2019, 144(8): 1941-1953.
- [15] Gutiérrez-Hoya A, Soto-Cruz I. NK cell regulation in cervical cancer and strategies for immunotherapy [J]. *Cells*, 2021, 10(11): 3104.
- [16] Jiang H. Latest research progress of liquid biopsy in tumor—a narrative review [J]. *Cancer Manag Res*, 2024, 16: 1031-1042.
- [17] 彭晓霞, 朱莉. 外泌体在宫颈癌中的研究与应用前景 [J]. *现代妇产科进展*, 2023, 32(3): 231-233.
Peng XX, Zhu L. Research and application prospects of exosomes in cervical cancer [J]. *Prog Obstetr Gynecol*, 2023, 32(3): 231-233.
- [18] Jiang W, Shi X, Sun L, et al. Exosomal miR-30a-5p promoted intrahepatic cholangiocarcinoma progression by increasing angiogenesis and vascular permeability in PDCD10 dependent manner [J]. *Int J Biol Sci*, 2023, 19(14): 4571-4587.
- [19] Lei L, Mou Q. Exosomal taurine up-regulated 1 promotes angiogenesis and endothelial cell proliferation in cervical cancer [J]. *Cancer Biol Ther*, 2020, 21(8): 717-725.
- [20] Xiang X, Poliakov A, Liu C, et al. Induction of myeloid-

- derived suppressor cells by tumor exosomes[J]. *Int J Cancer*, 2009, 124(11): 2621–2633.
- [20] Li H, Chi X, Li R, et al. HIV-1-infected cell-derived exosomes promote the growth and progression of cervical cancer[J]. *Int J Biol Sci*, 2019, 15(11): 2438–2447.
- [21] 杨翔, 陈焯, 郑玮, 等. 转移性宫颈癌细胞衍生的外泌体 miR-21-5p 促进宫颈癌侵袭及转移[J]. *新医学*, 2022, 53(12): 891–898.
- Yang X, Chen Y, Zheng W, et al. Exosomal miR-21-5p derived from metastatic cervical cancer cells promotes cervical cancer invasion and metastasis [J]. *J New Med*, 2022, 53(12): 891–898.
- [22] Luo H, Zhang H, Mao J, et al. Exosome-based nanoimmunotherapy targeting TAMs, a promising strategy for glioma[J]. *Cell Death Dis*, 2023, 14(4): 235.
- [23] Duan H, Liu Y, Gao Z, et al. Recent advances in drug delivery systems for targeting cancer stem cells [J]. *Acta Pharm Sin B*, 2021, 11(1): 55–70.
- [24] Liang Y, Duan L, Lu J, et al. Engineering exosomes for targeted drug delivery[J]. *Theranostics*, 2021, 11(7): 3183–3195.
- [25] Hao S, Bai O, Yuan J, et al. Dendritic cell-derived exosomes stimulate stronger CD8+ CTL responses and antitumor immunity than tumor cell-derived exosomes [J]. *Cell Mol Immunol*, 2006, 3(3): 205–211.
- [26] Hao S, Bai O, Li F, et al. Mature dendritic cells pulsed with exosomes stimulate efficient cytotoxic T-lymphocyte responses and antitumor immunity [J]. *Immunology*, 2007, 120(1): 90–102.
- [27] Zheng M, Hou L, Ma Y, et al. Exosomal let-7d-3p and miR-30d-5p as diagnostic biomarkers for non-invasive screening of cervical cancer and its precursors [J]. *Mol Cancer*, 2019, 18(1): 76.
- [28] Emam SE, Ando H, Abu Lila AS, et al. A Novel Strategy to increase the yield of exosomes (extracellular vesicles) for an expansion of basic research [J]. *Biol Pharm Bull*, 2018, 41(5): 733–742.
- [29] He L, Zhu D, Wang J, et al. A highly efficient method for isolating urinary exosomes[J]. *Int J Mol Med*, 2019, 43(1): 83–90.
- [30] Patel GK, Khan MA, Zubair H, et al. Comparative analysis of exosome isolation methods using culture supernatant for optimum yield, purity and downstream applications [J]. *Sci Rep*, 2019, 9(1): 5335.
- [31] 潘星昊, 刘明斌, 张晓燕, 等. 外泌体作为靶向治疗载体的研究进展[J]. *中国癌症杂志*, 2020, 30(9): 701–706.
- Pan XH, Liu MB, Zhang XY, et al. Research progress on exosomes as targeted therapy carriers [J]. *Chin Oncol*, 2020, 30(9): 701–706.
- [32] 吴忱, 辛林. 新的药物传递系统: 外泌体作为药物载体递送[J]. *中国生物工程杂志*, 2020, 40(9): 28–35.
- Wu Y, Xin L. New drug delivery systems: exosomes as drug carriers [J]. *J Chin Biotechnol*, 2020, 40(9): 28–35.
- [33] 窦沁怡, 王嘉政, 杨滢硕, 等. 外泌体来源非编码 RNA 在肿瘤微环境中的作用及临床应用价值[J]. *浙江大学学报(医学版)*, 2023, 52(4): 429–438.
- Dou QY, Wang JZ, Yang YS, et al. The role and clinical application value of exosome-derived non-coding RNAs in the tumor microenvironment [J]. *J Zhejiang Univ (Med Sci)*, 2023, 52(4): 429–438.
- [34] 范典, 郑博豪, 周圣涛. 宫颈癌靶向治疗和免疫治疗研究进展[J]. *中国肿瘤临床*, 2020, 47(21): 1100–1107.
- Fan D, Zheng BH, Zhou ST, et al. Progress in targeted therapy and immunotherapy for cervical cancer [J]. *Chin J Clin Oncol*, 2020, 47(21): 1100–1107.
- [35] Zhu X, Long L, Xiao H, et al. Cancer-derived exosomal miR-651 as a diagnostic marker restrains cisplatin resistance and directly targets ATG3 for cervical cancer [J]. *Dis Markers*, 2021, 2021: 1544784.
- [36] Konishi H, Hayashi M, Taniguchi K, et al. The therapeutic potential of exosomal miR-22 for cervical cancer radiotherapy [J]. *Cancer Biol Ther*, 2020, 21(12): 1128–1135.
- [37] Cho O, Kim DW, Cheong JY. Plasma exosomal miRNA levels after radiotherapy are associated with early progression and metastasis of cervical cancer: a pilot study [J]. *J Clin Med*, 2021, 10(10): 2110.
- [38] Tong Y, Jia L, Li M, et al. Identification of exosomal circSLC26A4 as a liquid biopsy marker for cervical cancer [J]. *PLoS One*, 2024, 19(6): e0305050.
- [39] 潘虹, 马可, 曹燕, 等. miR-30d-5p 对宫颈癌细胞生长和转移的影响[J]. *局解手术学杂志*, 2024, 33(8): 670–675.
- Pan H, Ma K, Cao Y, et al. Effect of miR-30d-5p on the growth and metastasis of cervical cancer cells [J]. *J Reg Anat Oper Surg*, 2024, 33(8): 670–675.
- [40] 王丽萍, 何婧, 许晓燕, 等. MiR-30a-5p 通过泛素水解酶 22 抑制人宫颈癌细胞上皮-间质转化功能[J]. *中山大学学报(医学版)*, 2019, 40(5): 698–705.
- Wang LP, He J, Xu XY, et al. MiR-30a-5p inhibits epithelial mesenchymal transition function in human cervical cancer cells by targeting ubiquitin-specific protease 22 [J]. *J Sun Yat-sen Univ (Med Sci)*, 2019, 40(5): 698–705.
- [41] Gu Y, Feng X, Jin Y, et al. Upregulation of miRNA-10a-5p promotes tumor progression in cervical cancer by suppressing UBE2L signaling [J]. *J Obstet Gynaecol*, 2023, 43(1): 2171283.
- [42] Liu X, Wang CX, Feng Q, et al. lncRNA TINCR promotes the development of cervical cancer via the miRNA-7/mTOR axis in vitro [J]. *Exp Ther Med*, 2023, 26(4): 487.
- [43] Aguilar-Martínez SY, Campos-Viguri GE, Medina-García SE, et al. MiR-21 regulates growth and migration of cervical

- cancer cells by RECK signaling pathway [J]. *Int J Mol Sci*, 2024, 25(7): 4086.
- [44] Srinath S, Jishnu PV, Varghese VK, et al. Regulation and tumor-suppressive function of the miR-379/miR-656 (C14MC) cluster in cervical cancer [J]. *Mol Oncol*, 2024, 18(6): 1608-1630.
- [45] Liu X, Liu Z, Liu Y, et al. ATG9A modulated by miR-195-5p can boost the malignant progression of cervical cancer cells [J]. *Epigenetics*, 2023, 18(1): 2257538.
- [46] He X, Sun J, Zhang J, et al. circ_0039787 promotes cervical cancer cell tumorigenesis by regulation of the miR-877-5p-KRAS axis [J]. *Aging (Albany NY)*, 2024, 16(3): 2736-2752.
- [47] Ni S, Wei Z, Li D. Effect of lncRNA LINC00324 on cervical cancer progression through down-regulation of miR-195-5p [J]. *J Obstet Gynaecol*, 2023, 43(2): 2285384.
- [48] Jin S, Wang W, Xu X, et al. miR-34b-3p-mediated regulation of STC2 and FN1 enhances chemosensitivity and inhibits proliferation in cervical cancer [J]. *Acta Biochim Biophys Sin (Shanghai)*, 2024, 56(5): 740-752.
- [49] Dabous E, Alalem M, Awad AM, et al. Regulation of KLRC and Ceacam gene expression by miR-141 supports cell proliferation and metastasis in cervical cancer cells [J]. *BMC Cancer*, 2024, 24(1): 1091.
- [50] Feng Y, Hang W, Sang Z, et al. Identification of exosomal and non-exosomal microRNAs associated with the drug resistance of ovarian cancer [J]. *Mol Med Rep*, 2019, 19(5): 3376-3392.
- [51] O'Malley DM, Krivak TC, Kabil N, et al. PARP inhibitors in ovarian cancer: a review [J]. *Target Oncol*, 2023, 18(4): 471-503.
- [52] Wang CK, Chen TJ, Tan GYT, et al. MEX3A mediates p53 degradation to suppress ferroptosis and facilitate ovarian cancer tumorigenesis [J]. *Cancer Res*, 2023, 83(2): 251-263.
- [53] Wu Y, Xu S, Cheng S, et al. Clinical application of PARP inhibitors in ovarian cancer: from molecular mechanisms to the current status [J]. *J Ovarian Res*, 2023, 16(1): 6.
- [54] Berek JS, Renz M, Kehoe S, et al. Cancer of the ovary, fallopian tube, and peritoneum: 2021 update [J]. *Int J Gynaecol Obstet*, 2021, 155 Suppl 1(Suppl 1): 61-85.
- [55] 吴若愚, 贾翔, 曹忆梦, 等. 外泌体在常见妇科恶性肿瘤诊治中的研究进展 [J]. *国际妇产科学杂志*, 2017, 44(3): 276-279.
- Wu RY, Jia X, Cao YM, et al. Research progress on exosome in the diagnosis and treatment of malignant gynecologic tumors [J]. *J Int Obstet Gynecol*, 2017, 44(3): 276-279.
- [56] 张杰克, 万少晖, 崔丽娟. 血清 AFP、CEA、CA19-9、CA125、HE4 水平联合检测在卵巢癌诊断中的应用价值 [J]. *中国民康医学*, 2023, 35(21): 113-116.
- Zhang JK, Wan SH, Cui LJ. Application value of combined detection of serum AFP, CEA, CA19-9, CA125 and HE4 levels in diagnosis of ovarian cancer [J]. *Med J Chin People's Health*, 2023, 35(21): 113-116.
- [57] 马心雨, 韩凤娟. 外泌体在卵巢癌中的作用机制及应用前景 [J]. *肿瘤预防与治疗*, 2024, 37(7): 547-554.
- Ma XY, Han FJ. Mechanism and application prospect of exosomes in ovarian cancer [J]. *J Canc Control Treat*, 2024, 37(7): 547-554.
- [58] Ho CM, Yen TL, Chang TH, et al. COL6A3 exosomes promote tumor dissemination and metastasis in epithelial ovarian cancer [J]. *Int J Mol Sci*, 2024, 25(15): 8121.
- [59] 周姣月, 刘洋, 周红林. 外泌体在妇科恶性肿瘤中的相关研究进展 [J]. *重庆医学*, 2020, 49(22): 3848-3851; +3854.
- Zhou JY, Liu Y, Zhou HL. Research progress of exosomes in gynecologic malignant tumors [J]. *Chongqing Med*, 2020, 49(22): 3848-3851; +3854.
- [60] 李洁, 姜洁. 卵巢癌靶向治疗应用及进展 [J]. *中国妇产科临床杂志*, 2024, 25(4): 289-291.
- Li J, Jiang J. Application and progress of targeted therapy for ovarian cancer [J]. *Chin J Clin Obstet Gynecol*, 2024, 25(4): 289-291.
- [61] Cao J, Wang H, Yang J, et al. Exosome-transmitted miR-30a-5p enhances cell proliferation, migration, and invasion in ovarian cancer [J]. *Cell Div*, 2023, 18(1): 20.
- [62] Wang H, Zhang Y, Miao H, et al. CircRAD23B promotes proliferation and carboplatin resistance in ovarian cancer cell lines and organoids [J]. *Cancer Cell Int*, 2024, 24(1): 42.
- [63] Liang H, Geng S, Wang Y, et al. Tumour-derived exosome SNHG17 induced by oestrogen contributes to ovarian cancer progression via the CCL13-CCR2-M2 macrophage axis [J]. *J Cell Mol Med*, 2024, 28(9): e18315.
- [64] Ran XM, Yang J, Wang ZY, et al. M2 macrophage-derived exosomal circTMC03 acts through miR-515-5p and ITGA8 to enhance malignancy in ovarian cancer [J]. *Commun Biol*, 2024, 7(1): 583.
- [65] Pei L, Zhao F, Zhang Y. USP43 impairs cisplatin sensitivity in epithelial ovarian cancer through HDAC2-dependent regulation of Wnt/ β -catenin signaling pathway [J]. *Apoptosis*, 2024, 29(1-2): 210-228.
- [66] Yang L, Yang Z, Liu Z, et al. Diagnostic value of plasma-derived exosomal miR-223 for epithelial ovarian cancer [J]. *BMC Womens Health*, 2024, 24(1): 150.
- [67] Sun L, Ke M, Yin M, et al. Extracellular vesicle-encapsulated microRNA-296-3p from cancer-associated fibroblasts promotes ovarian cancer development through regulation of the PTEN/AKT and SOCS6/STAT3 pathways [J]. *Cancer Sci*, 2024, 115(1): 155-169.

- [68] Liu J, Yan C, Xu S. LncRNA IL21-AS1 facilitates tumour progression by enhancing CD24-induced phagocytosis inhibition and tumorigenesis in ovarian cancer[J]. *Cell Death Dis*, 2024, 15(5): 313.
- [69] Liu S, Yuan L, Li J, et al. circDENND4C, a novel serum marker for epithelial ovarian cancer, acts as a tumor suppressor by downregulating miR-200b/c [J]. *Ann Med*, 2023, 55(1): 908-919.
- [70] Drouyer A, Beaussire L, Jorda P, et al. Clinical relevance of circulating ESR1 mutations during endocrine therapy for advanced hormone-dependent endometrial carcinoma [J]. *BMC Cancer*, 2023, 23(1): 1061.
- [71] Makker V, MacKay H, Ray-Coquard I, et al. Endometrial cancer[J]. *Nat Rev Dis Primers*, 2021, 7(1): 88.
- [72] Zhou L, Wang W, Wang F, et al. Plasma-derived exosomal miR-15a-5p as a promising diagnostic biomarker for early detection of endometrial carcinoma[J]. *Mol Cancer*, 2021, 20(1): 57.
- [73] Fan X, Cao M, Liu C, et al. Three plasma-based microRNAs as potent diagnostic biomarkers for endometrial cancer [J]. *Cancer Biomark*, 2021, 31(2): 127-138.
- [74] Fan X, Zou X, Liu C, et al. MicroRNA expression profile in serum reveals novel diagnostic biomarkers for endometrial cancer[J]. *Biosci Rep*, 2021, 41(6): BSR20210111.
- [75] Srivastava A, Moxley K, Ruskin R, et al. A non-invasive liquid biopsy screening of urine-derived exosomes for miRNAs as biomarkers in endometrial cancer patients [J]. *Aaps j*, 2018, 20(5): 82.
- [76] Shi S, Tan Q, Feng F, et al. Identification of core genes in the progression of endometrial cancer and cancer cell-derived exosomes by an integrative analysis [J]. *Sci Rep*, 2020, 10(1): 9862.
- [77] Jia J, Guo S, Zhang D, et al. Exosomal-lncRNA DLEU1 accelerates the proliferation, migration, and invasion of endometrial carcinoma cells by regulating microRNA-E2F3 [J]. *Onco Targets Ther*, 2020, 13: 8651-8663.
- [78] Wang J, Gong X, Yang L, et al. Loss of exosomal miR-26a-5p contributes to endometrial cancer lymphangiogenesis and lymphatic metastasis [J]. *Clin Transl Med*, 2022, 12(5): e846.
- [79] Gu X, Shi Y, Dong M, et al. Exosomal transfer of tumor-associated macrophage-derived hsa_circ_0001610 reduces radiosensitivity in endometrial cancer [J]. *Cell Death Dis*, 2021, 12(9): 818.
- [80] Fan JT, Zhou ZY, Luo YL, et al. Exosomal lncRNA NEAT1 from cancer-associated fibroblasts facilitates endometrial cancer progression via miR-26a/b-5p-mediated STAT3/YKL-40 signaling pathway [J]. *Neoplasia*, 2021, 23(7): 692-703.
- [81] Cai J, Wang R, Chen Y, et al. LncRNA FIRRE regulated endometrial cancer radiotherapy sensitivity via the miR-199b-5p/SIRT1/BECN1 axis-mediated autophagy [J]. *Genomics*, 2024, 116(1): 110750.
- [82] Ma H, Weng F, Tong X, et al. LncRNA TRPM2-AS promotes endometrial carcinoma progression and angiogenesis via targeting miR-497-5p/SPP1 axis [J]. *Cell Mol Biol Lett*, 2024, 29(1): 93.
- [83] Wang F, Zhou C, Zhu Y, et al. The microRNA Let-7 and its exosomal form: epigenetic regulators of gynecological cancers [J]. *Cell Biol Toxicol*, 2024, 40(1): 42.
- [84] 陆茜林, 丁娜, 贺红林, 等. 经血间充质干细胞外泌体对人卵巢癌细胞A2780生物学行为的影响[J]. *中国医学前沿杂志(电子版)*, 2024, 6(2): 50-57.
- Lu QL, Ding N, He HL, et al. Effects of exosomes derived from menstrual blood-derived mesenchymal stem cells on biological behavior of human ovarian cancer [J]. *Chin J Front Med Sci(Elect Ver)*, 2024, 16(2): 50-57.
- [85] Li R, Chibbar R, Xiang J. Novel EXO-T vaccine using polyclonal CD4 (+) T cells armed with HER2-specific exosomes for HER2-positive breast cancer [J]. *Onco Targets Ther*, 2018, 11: 7089-7093.
- [86] Zhang W, Zhou Q, Wei Y, et al. The exosome-mediated PI3k/Akt/mTOR signaling pathway in cervical cancer [J]. *Int J Clin Exp Pathol*, 2019, 12(7): 2474-2484.
- [87] Zhang X, Zhang H, Gu J, et al. Engineered extracellular vesicles for cancer therapy [J]. *Adv Mater*, 2021, 33(14): e2005709.
- [88] Naseri M, Bozorgmehr M, Zöller M, et al. Tumor-derived exosomes: the next generation of promising cell-free vaccines in cancer immunotherapy [J]. *Oncoimmunology*, 2020, 9(1): 1779991.

(编辑 黄子芸)